# **Retinoid x Receptor Alpha DNA Binding for Mitochondrial Transcription**

# James Weiser, Megan L. Bestwick Department of Chemistry, Linfield University, McMinnville, OR

#### Introduction

Mitochondria are important organelles in eukaryotic cells due to their role in metabolism. They also have their own genome (mtDNA) which encodes many of the protein subunits within the oxidative phosphorylation (OXPHOS) pathway. The remaining protein subunits are encoded in the nuclear genome, translated in the cytosol and imported into mitochondria. Transcriptional coordination between these two genomes to produce functional OXPHOS complexes is critical to maintaining cellular homeostasis.<sup>1</sup> A major control point is transcription, and this process requires regulatory proteins that determine the expression of these genes.<sup>1</sup> Retinoid X receptor alpha (RXRa) is a protein that has many functions in human cells and is a ligand activated nuclear receptor.<sup>2</sup> It can also translocate to the mitochondria where its purpose is unknown.<sup>3</sup> The goal of this project is to determine the role of RxRα as a mitochondrial transcription factor. We have expressed and purified the RXRa protein, and then also assessed its DNA binding affinity with various mtDNA sequences. We concluded RXRa binds as a dimer with space between the binding sites. Our aim is to investigate RxRa binding the mtDNA using atomic force microscopy (AFM). In doing so we will better understand the mechanisms that are involved in mtDNA transcription. If RxRa is a transcription factor that is another protein that we can regulate to increase or decrease transcription. Being able to increase or decrease transcription in mitochondria is important as mitochondrial dysfunction causes a wide array of diseases and plays a part in aging.

### Methods

Bacterial Transformation & Protein Expression

Plasmid DNA<sup>4</sup> (Figure 2) were extracted via a Zymo Research miniprep kit then transformed into to competent *E. coli* cells via heat shock. Cells were cultured in LB media for 2 days at 37°C with IPTG (0.1 mM) used to express the RxR $\alpha$  protein. Bacterial Lysis

Cells were pelleted by centrifugation. Buffer with Triton X-100 was added to the cells, vortexed and sonicated, then centrifuged again to separate out protein from other cell debris. RxRa Protein Purification

Protein was purified using GST affinity column chromatography (Cytiva) with PBS and then 10 mM glutathione to elute the protein. The protein was concentrated by filter centrifuging (Amicon). dsDNA Oligo Probe Formation

DNA oligo probes were created by combining the forward and reverse oligos, boiling, then letting it cool for 30 minutes to allow the dsDNA to anneal. The DR1 repeats were separated by 2 bases, and 5 bases (Figure 3), and third probe was tested with a single DR1 sequence (TGGTCA).<sup>5</sup>

DNA/Protein Complex

Electrophoretic mobility shift assay (EMSA) was used to assess the protein/DNA complex. Reactions included purified protein, BSA, 2x reaction buffer, DNA. Reactions were visualized on native gels.

## Results

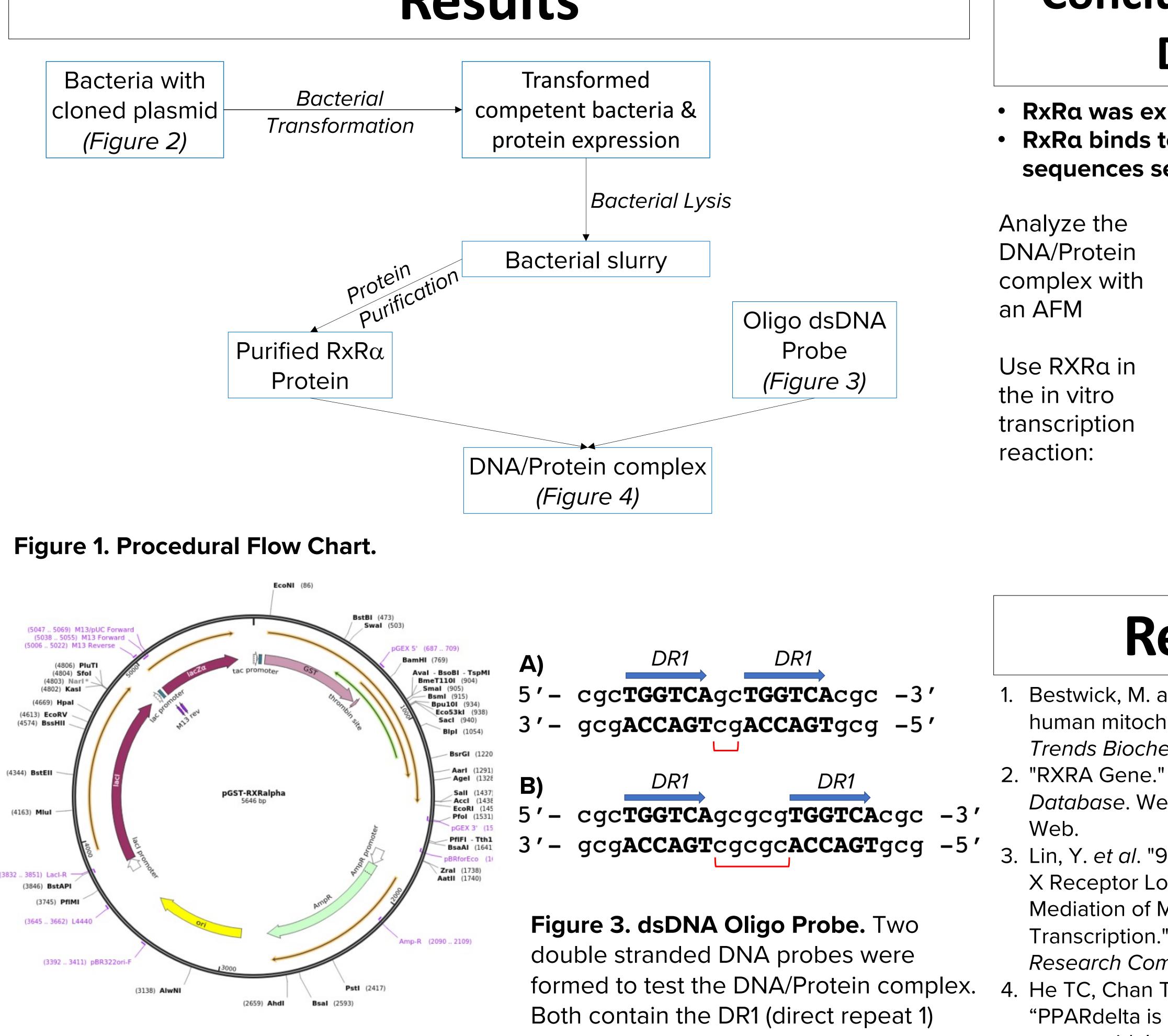
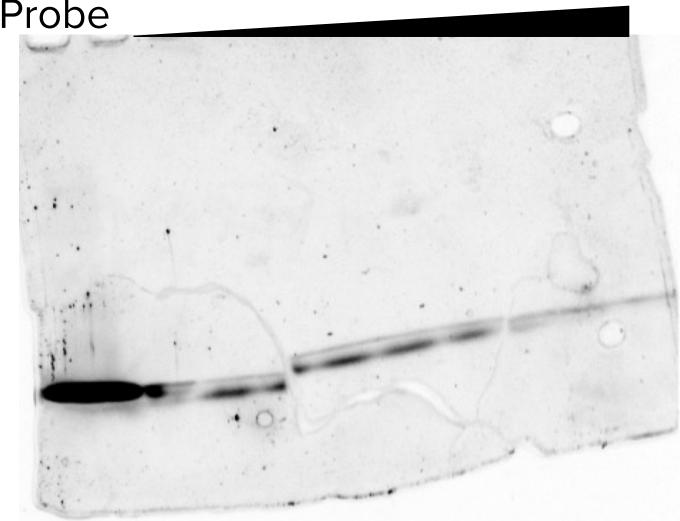


Figure 2. RxRα Expression Plasmid. pGST-RXRalpha was a gift from Bert Vogelstein (Addgene plasmid # 16547).

Figure 4. DNA/Protein Complex. EMSA was used to assess  $RxR\alpha$  binding to the DR1 repeats separated by 2 and 5 nucleotides. This image shows  $RxR\alpha$  binding to DR1 repeats separated by 5 nucleotides (2 ug probe). The RxR $\alpha$  concentration represents starting with 10 ug protein followed by a 1:5 serial dilution. Samples were run on a native acrylamide gel and visualize with GelRed.

Probe

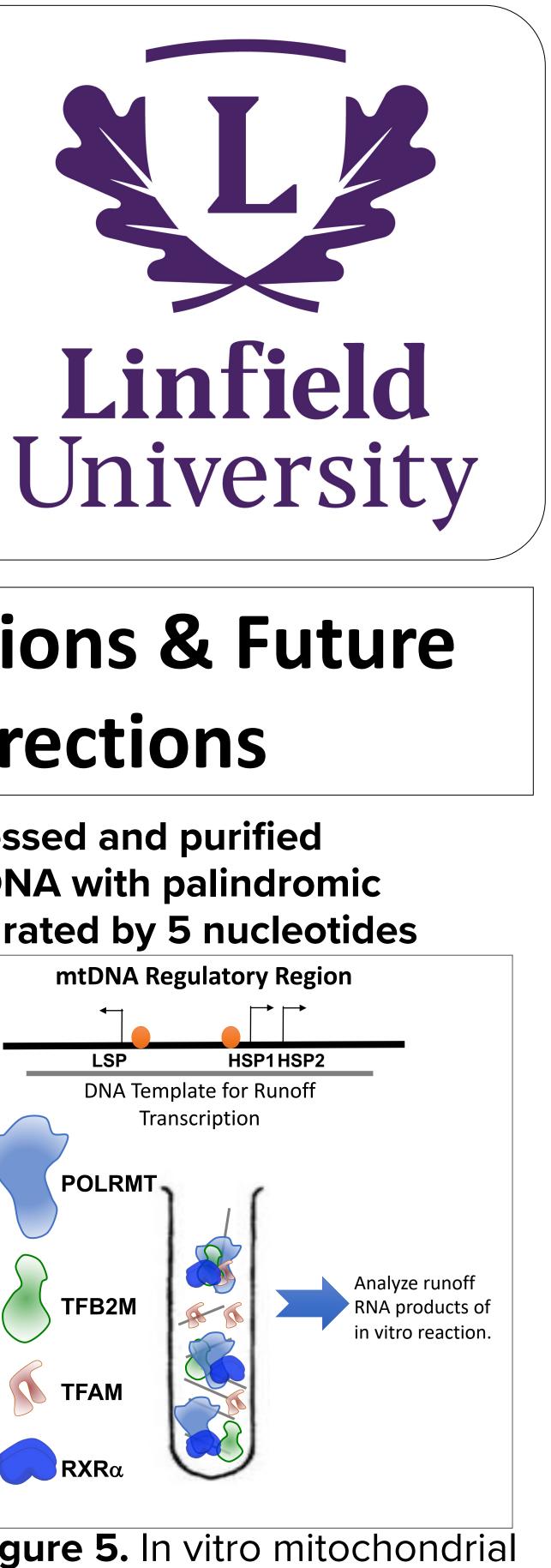


DR1	DR1	
<b>TGGTCA</b> gc <b>TGGTCA</b> cgc		-3′
g <b>ACCAGT</b> cg <b>ACCAGT</b> gcg		-5′

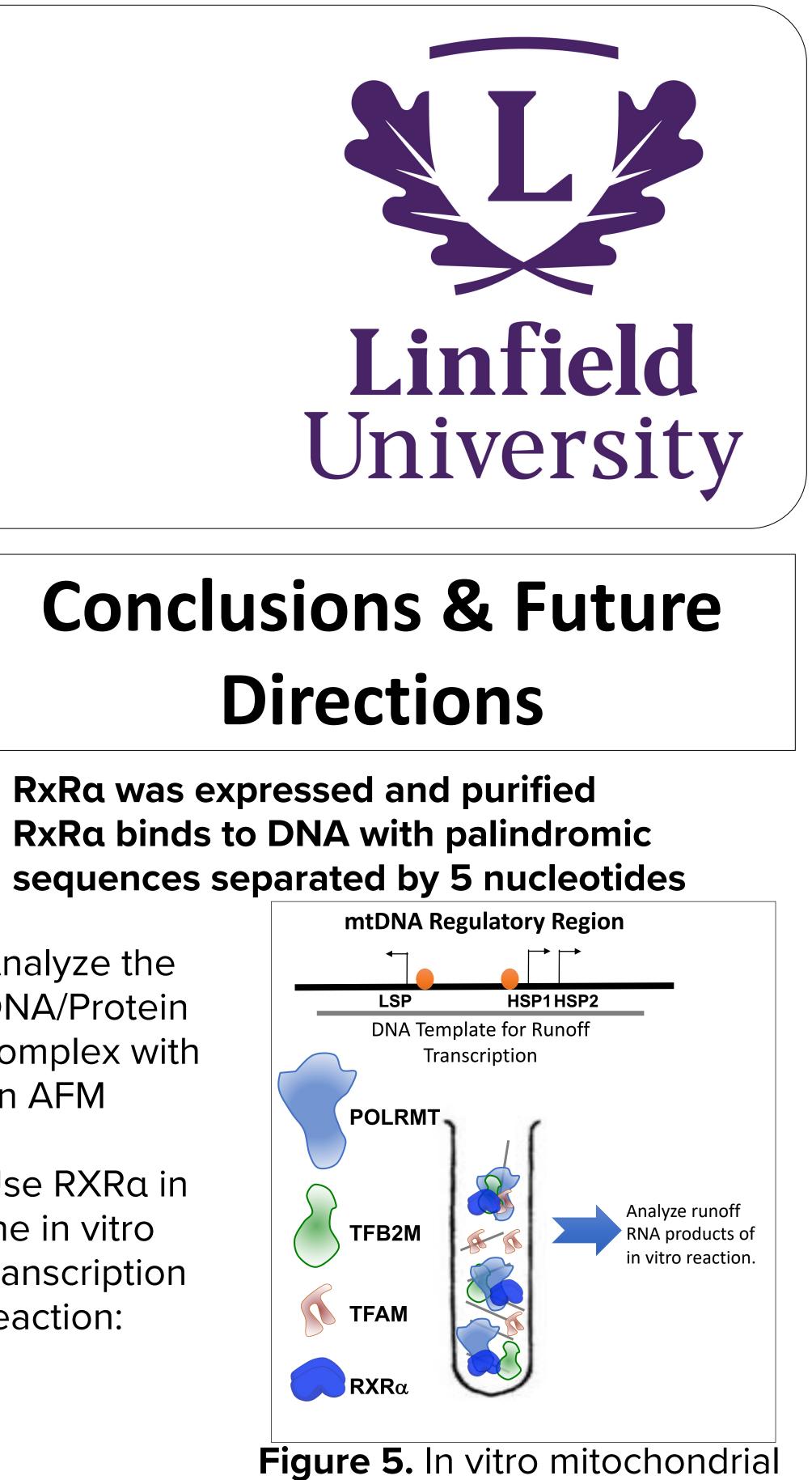
# sequcen in parallel. A) DR1 repeats separated by 2 nucleotides, B) DR1

repeats separated by 5 nucleotides.





- **RxRα** was expressed and purified



## References

- 2. "RXRA Gene." *Gene Cards Human Gene*
- Mediation of Mitochondrial Transcription." *Biochemical and Biophysical*
- 4. He TC, Chan TA, Vogelstein B, Kinzler KW. "PPARdelta is an APC-regulated target of (1999):335-45.
- 5. Rastinejad, F. et al. "Structure of the RXR-RAR DNA-binding complex on the retinoic acid 1045-54.

#### Acknowledgements

- Linfield Chemistry Department
- Linfield Biology & Physics Departments
- Dyke Science Endowment
- Linfield Research Institute
- M.J. Murdock Charitable Trust
- National Science Foundation

transcription assay.

Bestwick, M. and Shadel, G. S. "Accessorizing the human mitochondrial transcription machinery." Trends Biochemical Sciences 38:6 (2013): 283-91. Database. Weizmann Institute of Science, n.d.

3. Lin, Y. et al. "9-cis Retinoic Acid Induces Retinoid X Receptor Localized to the Mitochondria for *Research Communications* 377:2 (2008): 351-54. nonsteroidal anti-infammatory drugs." Cell 99:3

response element DR1." The EMBO J. 19:5 (2000):

Linfield University Collaborative Research Grant &