Fluorescent Detection of Reactive Oxygen Species in *Saccharomyces cerevisiae* Applied to Chronological Lifespan

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**Introduction**

During the course of normal aerobic metabolism, cells are exposed to a wide range of reactive oxygen species, such as the superoxide anion, hydrogen peroxide, the hydroxyl radical. These reactive oxygen species, or ROS, are highly reactive metabolites of oxygen and can damage a wide range of macromolecules in the cell (nucleic acids, proteins, and lipids) and can often times lead to cell death.

Normally, molecular oxygen is fairly unreactive and harmless in its’ ground state. However, it is able to undergo partial reduction from electrons leaked from the electron transport chain to form both the superoxide anion and hydrogen peroxide, both of which can react further to form the dangerously reactive hydroxyl radical. In order to combat the toxic and potentially deadly effects of ROS, cells are equipped with various antioxidant defense mechanisms. These mechanisms include enzymes like superoxide dismutase 1. When oxidative stress does occur, it is because a portion of ROS has overcome the host defenses like superoxide dismutase 1, resulting in oxidative damage to the cells.

It’s important to understand that superoxide dismutase 1 is primarily a cytosolic detoxifying enzyme that requires both copper and zinc cofactors to initiate and facilitate the conversion of the superoxide anion into hydrogen peroxide and then eventually, to water.

The overall objective of this research is to observe the abundance of different reactive oxygen species by applying various biochemical fluorescent staining assays to the chronological lifespan of the yeast, *Saccharomyces cerevisiae*.

**Materials and Methods**

- *S. cerevisiae* of superoxide dismutase 1 deletion strain (ΔSOD1), which was expected to have ROS sensitivity and a Wild Type (genetically the same, but no SOD1 deletion), which was expected to have ROS resistance. Both were of the BY4743 strains.
- Cultures were inoculated (OD<sub>600</sub> = 0.5) in full YPD media and grown for 24 hrs. Cultures (1 mL) were added to minimal media (49 mL) and grown for 24 hrs.
- Experimental cultures were treated (20 min, 37 C) with 100 µL DHE+PBS stock solution (5 µL DHE, 995 µL PBS).
- Shimadzu RF-6000 scanning spectrofluorimeter was used to measure parameters of fluorescence at an emission range of 300-700 nm and an excitation of 405 nm and SpectraMax Gemini XPS was used to measure parameters of fluorescence at an emission of 325 nm and an excitation of 461 nm.
- Dihydroethidium (DHE) was the biochemical dye used to quantitate levels of different reactive oxygen species produced by the yeast strains because it was the most specific and least problematic, as it is retained well by *S. cerevisiae*.

**Results and Conclusions**

Table 1 shows the intensity of fluorescence when the excitation was 405 nm and when the emission was 470 nm. There is a distinguishable difference in observable fluorescence between the WT strain treated with DHE and the WT strain not treated with DHE. However, we will need to do more trials to determine if this is a significant difference that can be reproduced. Values were normalized to the untreated WT strain, so that results could be more easily compared. When looking to the ΔSOD1 strain, however, our results were not what we predicted. As exemplified by Table 1, there was more fluorescence observed overall in the wild type than in the ΔSOD1/with DHE. It would be expected that a yeast strain deleted for SOD1 would display more reactive oxygen species due to the lack of the detoxifying enzyme, but here we see greater fluorescence in the WT strains (indicating more ROS). It is clear that this data must be developed and reproduced further so that it can be understood as to why more ROS was being produced in the WT strain than in the ΔSOD1 strain.

The results from the Shimadzu RF-6000 scanning spectrofluorimeter, provided us with solid preliminary data about DHE and how it reacts with Wild Type yeast. Even though there was some background in the trials, we can compare the Wild Type yeast strains quite well and can see that there is a greater intensity of fluorescence in the strain treated with DHE (Fig. 2) compared to the strain that wasn’t. Hence, there is reason to believe that reactive oxygen species was able to be tracked using DHE. Figure 1 illustrates the published spectra for DHE.

Our further exploration of fluorescence detection with the SpectraMax Gemini XPS confirmed that we were able to track detection of reactive oxygen species in both strains and also eliminate the background noise present in our previous experiment (see Figure 2).

**Future Directions**

- Future research will focus on obtaining more replicable results and establishing a greater difference between fluorescence emitted from the WT yeast with and without DHE.
- The discrepancies in the ROS presence in the ΔSOD1 strains will also be further investigated by integrating the use of various positive controls such as respiratory chain inhibitors (Antimycin A and Oligomycin) in order to obtain replicable and distinguished results that will allow for the acquisition of a statistically significant, quantifiable difference in fluorescence between WT and ΔSOD1 cultures.
- Capture images via microscopy of reactive oxygen species in live yeast at various points during chronological lifespan.

**Literature Cited**


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**Further Information**

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